

Sensors Integration in Organs-on-Chip

The integration of sensors in organs-on-chip platforms to monitor cellular processes is challenging. Flow conditions and geometries must be optimized to improve detection.

J. Charmet¹, S. Wenger¹, A. Tekari¹, S. Schmidt², L. Burr³, L. Suter-Dick², A. Homsy¹

¹ Haute Ecole Arc, HES-SO University of Applied Sciences and Arts of Western Switzerland, Neuchâtel, Switzerland

² FHNW, Institute for Chemistry and Bioanalytics, Muttenz, Switzerland

³ CSEM, Landquart, Switzerland

Abstract

Organs-on-Chips (OoCs) are microfluidics-based systems that support complex cell cultures systems in controllable physiological conditions¹. Such systems are expected to reduce animal testing and accelerate drug discovery². OoCs can be interfaced with biosensors for real-time and high-throughput

measurements. However, such an integration is challenging. Biosensors can rarely operate continuously without degradation of performance whereas OoC operation can last several days or weeks. In addition, changes in molecular concentrations in OoCs can be extremely low.

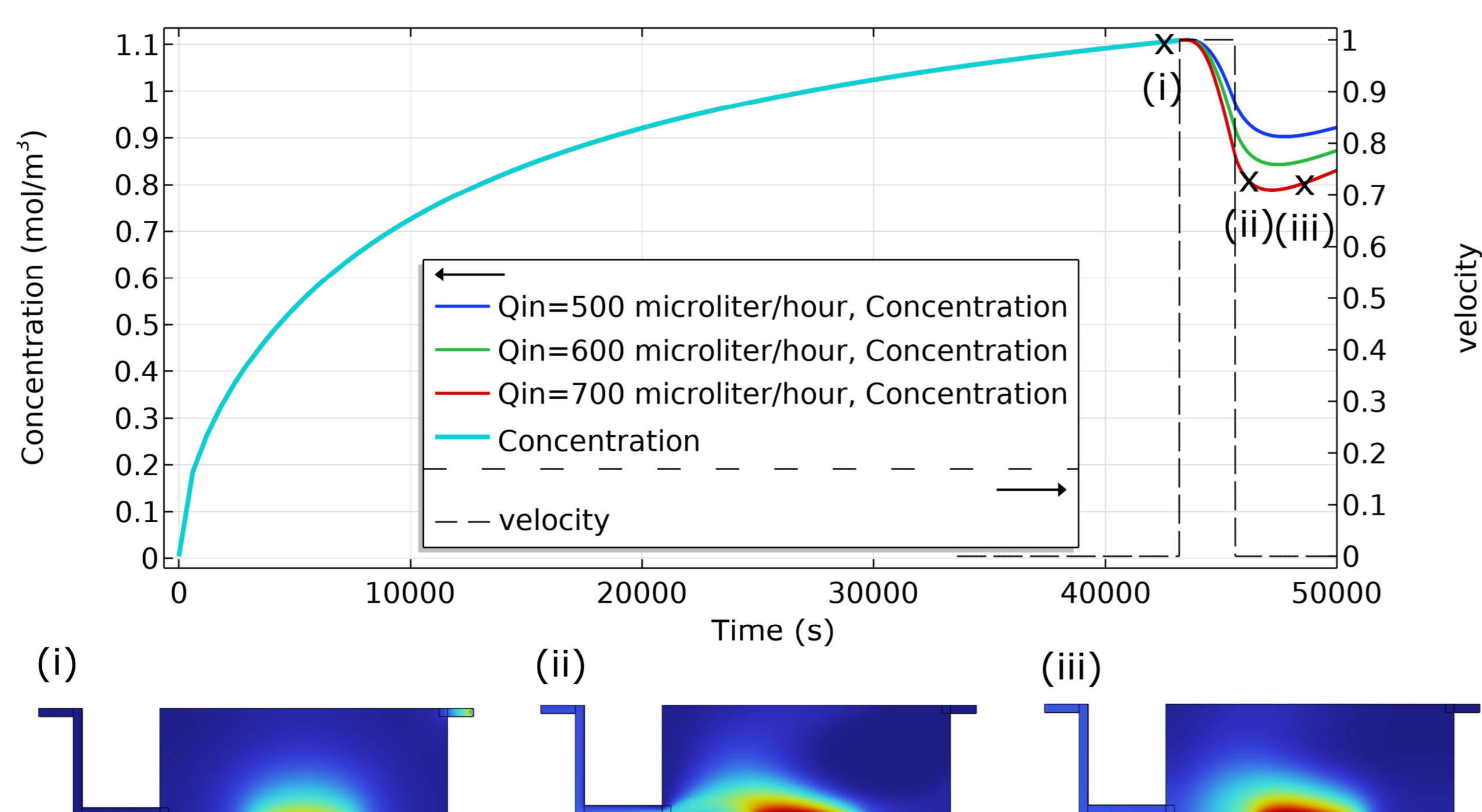


Figure 1. Concentration in the chamber as a function of time and flow rates with evolution of the concentration profile in the chamber at different times.

Results

Using a first model (not shown), we demonstrated that an incubation time (without flow) was necessary to accumulate enough molecular concentration changes in the chamber to reach the limit of detection of the sensor.

The flow conditions (delay, flow rate and duration) were systematically studied to optimise the transfer of lactate to the sensors. Figure 2 shows that high flow rates may result in excessive dilution.

Methodology

To optimise the flow conditions and dimensions of the platform, we use the Laminar Flow, Transport of Diluted Species and General Form Boundary PDE physics interfaces. The 3D symmetric model consists of a two-step study (Figure 1) to account for the incubation (hours timescale) and the active transport (minutes timescale) steps. Active transport is defined by a rectangle function (dashed line). A parametric sweep in the second study enables the evaluation of several flow and molecular conditions.

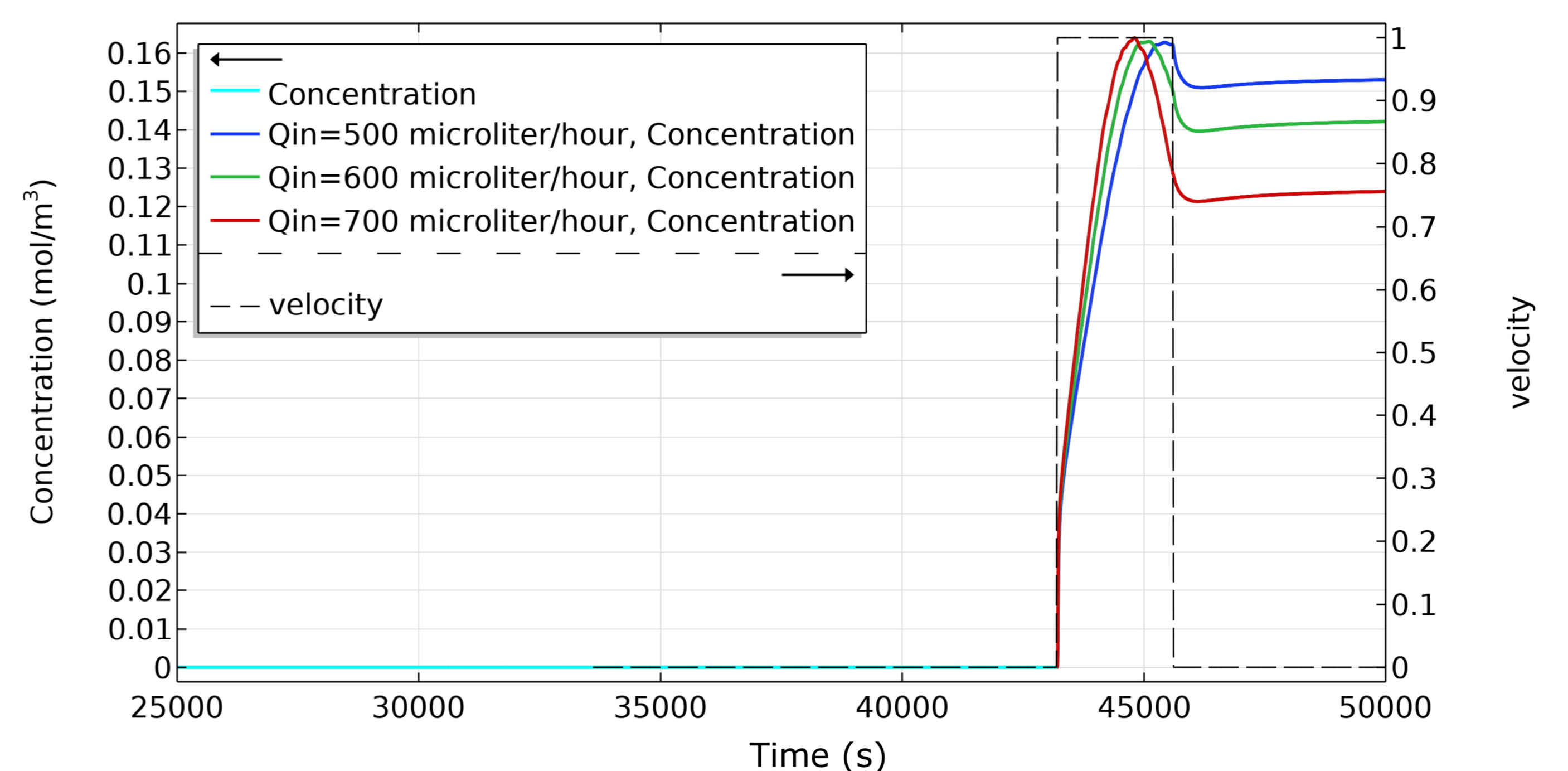


Figure 2. Concentration at the sensor area for different flow rates (500 $\mu\text{L/h}$, 600 $\mu\text{L/h}$ and 700 $\mu\text{L/h}$) after 12 hours of incubation and 40 minutes of active transport.

REFERENCES

[1] Wang, K.; Man, K.; Liu, J.; Liu, Y.; Chen, Q.; Zhou, Y.; Yang, Y. Microphysiological Systems: Design, Fabrication, and Applications. *ACS Biomater. Sci. Eng.* 2020, 6 (6), 3231.

[2] Loewa, A.; Feng, J. J.; Hedtrich, S. Human Disease Models in Drug Development. *Nat. Rev. Bioeng.* 2023, 1 (8), 545–559.

haute école **arc** ingénierie
neuchâtel berne jura www.he-arc.ch

csem

n|w

University of Applied Sciences and Arts
Northwestern Switzerland